

The Neural Coding of Feedback Learning across Child and Adolescent Development

Sabine Peters^{1,2}, Barbara R. Braams^{1,2}, Maartje E. J. Raijmakers³,
P. Cédric M. P. Koolschijn^{1,2,3*}, and Eveline A. Crone^{1,2,3*}

Abstract

■ The ability to learn from environmental cues is an important contributor to successful performance in a variety of settings, including school. Despite the progress in unraveling the neural correlates of cognitive control in childhood and adolescence, relatively little is known about how these brain regions contribute to learning. In this study, 268 participants aged 8–25 years performed a rule-learning task with performance feedback in a 3T MRI scanner. We examined the development of the frontoparietal network during feedback learning by exploring contributions of age and pubertal development. The pFC showed more activation following negative compared with positive feedback with increasing age. In contrast, our data suggested

that the parietal cortex demonstrated a shift from sensitivity to positive feedback in young children to negative feedback in adolescents and adults. These findings were interpreted in terms of separable contributions of the frontoparietal network in childhood to more integrated functions in adulthood. Puberty (testosterone, estradiol, and self-report) did not explain additional variance in neural activation patterns above age, suggesting that development of the frontoparietal network occurs relatively independently from hormonal development. This study presents novel insights into the development of learning, moving beyond a simple frontoparietal immaturity hypothesis. ■

INTRODUCTION

A crucial aspect of successful learning is the ability to process performance feedback and to adjust behavior on subsequent occasions, also referred to as feedback learning (Holroyd & Coles, 2002). Across development, children show marked improvements in feedback learning (Eppinger, Mock, & Kray, 2009; Crone, Wendelken, Donohue, & Bunge, 2006; Welsh, Pennington, & Girois, 1991), which is evident from performance on neuropsychological tasks such as the WCST and experimental switch tasks such as task-switching paradigms (Eppinger et al., 2009; van den Bos, Guroglu, van den Bulk, Rombouts, & Crone, 2009). Given the importance of learning in school settings, it is essential to unravel the mechanisms behind the development of feedback learning.

Prior studies of brain mechanisms for negative feedback processing in adults showed increased activation in dorsolateral pFC (DLPFC), pre-SMA/ACC, and superior parietal cortex (SPC; Zanolie, Van Leijenhorst, Rombouts, & Crone, 2008) after receiving negative feedback relative to positive feedback. In children, these areas showed less activation following negative feedback, relative to positive feedback, compared with adults (Crone, Zanolie,

Van Leijenhorst, Westenberg, & Rombouts, 2008). This finding is consistent with evidence that these regions are still developing during adolescence, in terms of both structure (Raznahan et al., 2011; Shaw et al., 2008) and function (Klingberg, Forssberg, & Westerberg, 2002; Tamm, Menon, & Reiss, 2002).

Until recently, most theories on the development of cognitive control were based on the assumption that, with age, the frontoparietal network comes increasingly “online” in a linear fashion (Somerville, Jones, & Casey, 2010; Bunge & Wright, 2007). However, recent reviews questioned whether cognitive development is associated with a linear increase in the frontoparietal network (Crone & Dahl, 2012; Pfeifer & Allen, 2012). For example, prior studies on feedback learning indicated that children showed not only less activation after negative feedback compared with adults but also more activation in the same regions in the frontoparietal network following positive feedback (van den Bos et al., 2009; van Duijvenvoorde, Zanolie, Rombouts, Raijmakers, & Crone, 2008). This suggests that there may be a qualitative shift with development in recruitment of brain areas for feedback learning.

The interpretation of prior studies is complicated because of differences in age group selection. Some studies compared 8-year-olds with 12-year-olds (van Duijvenvoorde et al., 2008), whereas others collapsed across 8- to 12-year-olds (Eppinger et al., 2009; Crone et al., 2008).

¹Leiden University, ²Leiden Institute for Brain and Cognition,
³University of Amsterdam

*Shared last author.

Adolescent groups were selected from a wide age range from 13 to 16 years, which is problematic given that neural development continues in this period (van den Bos et al., 2009). Additionally, prior studies have not investigated the influence of pubertal maturation, which may explain additional variance in developmental change (Forbes & Dahl, 2010). For example, it was previously found that self-reported puberty scores explained additional variance over age in neural activity in a working memory study (Schweinsburg, Nagel, & Tapert, 2005). Others have suggested that brain regions supporting cognitive control develop relatively independent of pubertal influences (Steinberg et al., 2008). A study including participants across the whole range of childhood and adolescence, focusing on both age and puberty effects, has not yet been performed.

In this study, we tested 268 participants between the ages of 8 and 25 to pinpoint developmental differences in feedback learning with increased precision. Participants performed a learning task in which stimuli were sorted at one of three possible locations and choices were followed by negative or positive feedback. To control for informative value of feedback, we distinguished between a learning phase and an application phase. These phases were based on a distinction between early feedback (informative for learning) and late feedback (uninformative for learning; Eliassen et al., 2012). We formed two hypotheses: (a) We predicted more activation in DLPFC, pre-SMA/ACC, and SPC after feedback, both negative and positive, during learning compared with application of rules. We hypothesized that distinguishing between feedback informative for learning (learning phase) and uninformative feedback (application phase) is an ability that develops with age and contributes to performance. (b) We predicted more activity in adults than in children in DLPFC and SPC after negative feedback and more activity in children than in adults in DLPFC and SPC after positive feedback (van Duijvenvoorde et al., 2008). In addition, we tested at what age activity patterns become adult-like and whether the transition between childhood and adulthood can be explained with both age and puberty as predictors (Casey, Jones, & Somerville, 2011).

METHODS

Participants

The final sample (after exclusions) included 268 participants (138 female) between 8.01 and 25.95 years old ($M = 14.22$, $SD = 3.63$), who were recruited through local schools and advertisements. We selected age groups such that each age was represented by $n > 20$. Because of the relatively small number of 8- and 9-year-olds, they were combined into one age group to ensure similar group sizes. See Table 1 for the final number of participants per age group and per sex. A chi-square test indicated that the proportion of males to females was similar across age

Table 1. Number of Participants per Sex and per Age Group

Age (years)	Female	Male	Total
8/9	20	9	29
10	11	12	23
11	13	14	27
12	19	11	30
13	16	20	36
14	10	17	27
15	10	11	21
16	11	9	20
17	12	11	23
18–25	16	16	32
Total	138	130	268

groups ($\chi^2(9) = 8.70$, $p = .465$). IQ was estimated with two subtests of the WAIS-III or WISC-III (Similarities and Block Design). All estimated IQ scores were within the normal range ($M = 110.25$, $SD = 10.62$, range = 80–143). Adults received payment for participation, and children and their parents received a present and travel reimbursement. None of the participants reported a history of neurological or psychiatric disorders or current use of psychotropic medication. This study was approved by the internal review board at the Leiden University Medical Center, and all participants (or participant's parents in the case of minors) provided written informed consent. All anatomical MRI scans were reviewed and cleared by a radiologist.

Exclusion Criteria

Twenty-five participants were excluded from the analyses for the following reasons: Nineteen were excluded because movement in the MRI scanner exceeded 3.0 mm. Three participants were excluded because of damaged fMRI scans. Finally, three participants were excluded because they were extreme outliers on the total percentage of positive feedback (more than three times the interquartile range), thus indicating that they did not perform the task adequately.

Feedback Learning Task

Participants performed a child-friendly feedback learning task in the MRI scanner. On each trial, they saw a screen with three empty squares, under which a stimulus was presented (one of three possible stimuli; see Figure 1). The participants were told that each stimulus belonged in one of the three squares and their task was to sort the stimuli into the correct square. Performance feedback

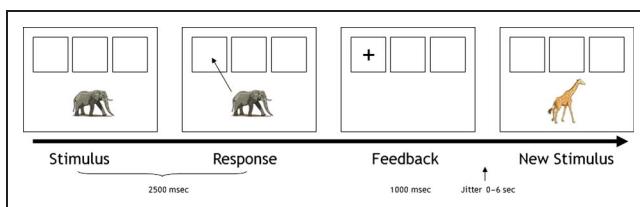


Figure 1. Display of task sequence.

was a plus sign (“+”) for a correct sort (positive feedback) and a minus sign (“−”) for an incorrect sort (negative feedback). The stimuli were presented in a pseudorandom order, with a maximum of two identical stimuli in a row. After 12 trials or when the participant applied the correct location twice in total for each stimulus, the sequence ended and a new sequence was presented with three new stimuli. There were 15 sequences in total, resulting in a maximum of 180 trials. Before the MRI session, all participants practiced three sequences. The task was divided into two runs of eight and seven sequences, respectively. Each trial started with a 500-msec fixation cross. Stimuli were presented for 2500 msec during which the response had to be given. Next, feedback was presented for 1000 msec. Intertrial intervals were jittered based on OptSeq (Dale, 1999), with intervals varying between 0 and 6 sec. The 500-msec fixation cross was presented for all trials and was not part of the ISI.

Feedback Types

For each sequence, we made a distinction between a learning phase and an application phase. The learning phase was defined as those trials where participants had not yet found the correct location for the stimulus and were using trial-and-error or hypothesis testing to find the correct solution. The application phase was defined as those trials where a stimulus was sorted correctly on a preceding trial and which continued to be sorted correctly on subsequent trials. Some trials in the learning phase did not actually result in learning. That is, the feedback on these stimuli was not used correctly in a subsequent trial. These trials were excluded from the analysis ($M = 3.65\%$, $SD = 3.03\%$ of all trials). Taken together, we defined the following three feedback types:

Learning phase. (a) PositiveLearning: refers to the sequence [CORRECT, correct] trials (upper case refers to the current trial, lower case refers to the preceding or subsequent stimulus): A first encountered correct feedback of a stimulus followed by a correct sort on the next trial of this stimulus. (b) NegativeLearning: refers to the sequence [ERROR, correct or error] trials: A first encountered error feedback of a stimulus followed by a choice for another location on the next trial of this stimulus.

Application phase. (c) Application: refers to the sequence [correct, CORRECT] trials: A correct feedback of a stimulus preceded by a correct sort on an earlier trial for that stimulus.

Learning Performance

To measure performance, we calculated the “learning rate” for each participant. This was defined as the percentage of trials in the learning phase where feedback was successfully used on the next trial. For this purpose, we divided the number of trials scored as PositiveLearning or NegativeLearning by the total number of trials during the learning phase.

Pubertal Development Measures

Pubertal Development Scale

To assess pubertal status, the Pubertal Development Scale (PDS; Petersen, Crockett, Richards, & Boxer, 1988) was completed by participants under 18 (participants from 18–25 years were given the maximum score for pubertal development). Physical development was reported on five questions on a 4-point scale. Girls reported body growth, body hair, breast development, skin changes, and menarche; boys reported body growth, body hair, facial hair, skin changes, and voice changes. Fourteen girls and seven boys as well as the adults (18–25 years) did not fill out the PDS list, leaving the total number of included PDS scores at 220. Total PDS score was calculated as the mean of the five questions. Mean PDS score was 2.55 for girls ($SD = 0.91$, range 1.00–4.00), 2.29 for boys ($SD = 0.82$, range 1.00–4.00), and 2.42 for boys and girls combined ($SD = 0.87$, range 1.00–4.00).

Sex Steroid Levels

In addition to self-report measures of puberty, we collected saliva measures to extract testosterone and estradiol levels (de Water, Braams, Crone, & Peper, 2013; Peper et al., 2012). Boys and girls collected saliva by passive drool at home, directly after waking up. Girls using contraceptives collected saliva on the last day of the stopping period (Day 7) and postmenarchal girls collected saliva on the seventh day of the menstrual cycle to control for hormonal fluctuations during the cycle. Girls using contraceptives without a stopping period, such as hormonal intrauterine devices, were excluded from this study. The saliva samples were assayed for testosterone and estradiol levels at the Department of Clinical Chemistry of the Free University Medical Centre. The lower detection limit was 4 pmol/L for testosterone and 0.1 pg/ml for estradiol.

Testosterone levels from saliva were determined by isotope dilution—online solid phase extraction liquid chromatography-tandem mass spectrometry (ID-XLC-MS/MS; Peper et al., 2012). Intraassay coefficient of variation (CV) was 11% and 4% at 10 and 140 pmol/L, respectively,

and interassay CV was 8% and 5% at 31 and 195 pmol/L, respectively (de Water et al., 2013). Seventeen girls and 14 boys did not succeed in collecting an appropriate quantity of saliva for analysis, leaving the total number of included hormonal samples at 237. Testosterone levels were highly skewed; thus, a log transformation of the scores was used for further calculations. Testosterone levels were highly correlated with self-report PDS in both boys ($r = .70$, $p < .001$) and girls ($r = .64$, $p < .001$). Salivary estradiol was determined using an enzyme-linked immunosorbent assay (DRG Instruments, Marburg, Germany). Interassay CV was 8% and 15% at 10 and 40 pg/L, respectively. For 11 girls and 15 boys, estradiol could not be determined, leaving the number of included samples at 242. Estradiol levels were correlated with PDS in both boys ($r = .23$, $p = .013$) and girls ($r = .24$, $p = .007$).

MRI Data Acquisition

MRI scans were acquired with a standard whole-head coil on a Philips 3.0T MRI scanner. Functional scans were acquired during two runs with T2*-weighted EPI. The first two volumes were discarded to allow for equilibration of T1 saturation effects. Volumes covered the whole brain (repetition time = 2.2 sec, echo time = 30 msec, sequential acquisition, 38 slices, slice thickness = 2.75 mm, field of view = $220 \times 220 \times 114.68$ mm). A high-resolution 3-D T1-FFE scan for anatomical reference was obtained after the experimental tasks (repetition time = 9.76 msec, echo time = 4.59 msec, 140 slices, voxel size = 0.875 mm, field of view = $224 \times 177 \times 168$ mm). The experimental task was projected on a screen that was viewed through a mirror. Before the MRI session, participants were accustomed to the MRI environment and sounds with a mock scanner.

fMRI Data Analysis

All data were analyzed with SPM8 (Wellcome Trust Centre for Neuroimaging, London). Images were corrected for slice timing acquisition and rigid body motion. Structural and functional volumes were spatially normalized to T1 templates. The normalization algorithm used a 12-parameter affine transform together with a nonlinear transformation involving cosine basis functions and resampled the volumes to 3-mm cubic voxels. Templates were based on the MNI305 stereotaxic space (Cocosco, Kollokian, Kwan, & Evans, 1997), an approximation of Talairach space (Talairach & Tournoux, 1988). Functional volumes were spatially smoothed with an 8-mm FWHM isotropic Gaussian kernel. The fMRI time series data were modeled by a series of events convolved with a canonical hemodynamic response function. The modeled events were “PositiveLearning” (sequence [CORRECT, correct], “NegativeLearning” (sequence [ERROR, correct or error]) and “Application” (sequence [correct, CORRECT]), which were time-locked to the moment of feedback. Other

trials including trials where participants responded too late were modeled separately but were not included in the analysis (events of no interest). The events (trials) were used as covariates in a general linear model, along with a basic set of cosine functions that high-pass filtered the data. The least squares parameter estimates of height of the best-fitting canonical hemodynamic response function for each condition were used in pairwise contrasts. The resulting contrast images, computed on a subject-by-subject basis, were submitted to higher-level analyses, also referred to as group analyses. We calculated the contrast Learning (positive and negative feedback combined) > Application to examine which areas contribute to learning from positive and negative feedback. To investigate valence effects, the contrasts PositiveLearning > NegativeLearning and NegativeLearning > PositiveLearning were calculated. We used a 10-voxel spatial extent combined with a stringent FWE-corrected intensity threshold to produce a desirable balance between Types I and II error rates (Bennett, Wolford, & Miller, 2009; Forman et al., 1995).

ROI Analysis

To examine transitions in age effects in more detail, ROI analyses were performed with the Marsbar toolbox in SPM8 (Brett, Anton, Valabregue, & Poline, 2002). The contrast used to generate functional ROIs was Learning (positive and negative) > Application (FWE corrected, $p < .05$, >10 contiguous voxels). We chose this contrast, because it is not biased toward positive or negative feedback but at the same time reveals task-relevant areas. The resulting ROIs spanned several brain regions. Therefore, the ROIs were subdivided by masking the functional ROI with the following anatomical Marsbar ROIs (based on automated anatomical labeling): left and right DLPFC (middle frontal gyrus), pre-SMA/ACC (SMA; left and right combined), left and right SPC (superior parietal gyrus). These ROIs were selected based on prior research indicating that these areas show age differences for feedback learning (Crone et al., 2008; van Duijvenvoorde et al., 2008). The DLPFC ROIs were very large (right: 28,488 mm; left: 28,240 mm), therefore, we created 6-mm radius spheres based on four local maxima within the DLPFC regions (two per hemisphere). These areas are referred to as “superior DLPFC” and “mid-DLPFC”. Center-of-mass MNI (x y z) coordinates were as follows: pre-SMA/ACC: 0 9 58; right superior DLPFC: 21 9 57; left superior DLPFC: -24 3 57; right mid-DLPFC: 42 18 39; left mid-DLPFC: -42 24 39; right SPC: 27 -62 55; left SPC: -23 -64 50.

Structural MRI Analysis

Cortical reconstruction was measured automatically using FreeSurfer version 5.0 (surfer.nmr.mgh.harvard.edu/; Fischl & Dale, 2000; Dale, Fischl, & Sereno, 1999; Fischl, Sereno, & Dale, 1999). Details of the surface-based cortical reconstruction have been documented in detail previously

(Fischl, Liu, & Dale, 2001; Fischl & Dale, 2000; Dale et al., 1999; Fischl, Sereno, & Dale, 1999; Fischl, Sereno, Tootell, & Dale, 1999). To extract cortical thickness from the functional ROIs, we performed the following steps: (1) Each functional ROI (bilateral mid and superior DLPFC, pre-SMA/ACC, bilateral SPC) was registered automatically to the FreeSurfer “fsaverage” template and inspected for accuracy of registration. Of note, as FreeSurfer calculates cortical thickness per hemisphere, the pre-SMA/ACC ROI was split into a left and right structural ROI. (2) Individual cortical thickness data were mapped to the “fsaverage” template. (3) Cortical thickness in millimeters was extracted for each ROI and individual separately. (4) For functional analyses, the bilateral pre-SMA/ACC cortical thickness ROI was averaged back to one ROI. We used an average weighted procedure by taking into account hemispherical differences in surface size maps. In all areas, cortical thickness decreased with age (pre-SMA/ACC: $r = -.40$, $p < .001$; right superior DLPFC: $r = -.15$, $p = .022$; left superior DLPFC: $r = -.21$, $p = .002$, right mid-DLPFC: $r = -.19$, $p = .004$; left mid-DLPFC: $r = -.29$, $p < .001$; right SPC: $r = -.27$, $p < .001$; left SPC: $r = -.27$, $p < .001$).

Statistical Analyses

Behavioral and ROI fMRI data were analyzed with SPSS 19 (IBM Corp., Armonk, NY). Age effects were investigated by calculating Pearson’s correlations between age and several measures of interest. For the behavioral and ROI analyses, Age was used as a categorical variable in ANOVAs, divided in 10 age groups (8/9, 10, 11, 12, 13, 14, 15, 16, 17, 18–25). We reasoned that this approach allows for a precise index of age effects and allows for comparison with prior literature. Least significant difference (LSD) post hoc tests were performed to further investigate significant results. Hierarchical linear regression analyses were performed with SPSS 19 to investigate contributions of age and puberty to performance and neural measures. Age and puberty were added as continuous variables in regression analyses, because this provides the best method to test the relative contributions of both developmental indices. Finally, the contributions of cortical thickness to ROI activity were investigated with a hierarchical linear regression with age and cortical thickness as predictors.

RESULTS

Behavior

On average, participants needed 138.66 trials ($SD = 9.11$, range = 117–165) to complete the task. In general, participants had a high learning rate ($M = 93.39\%$, $SD = 5.11\%$). A t test indicated no sex differences for learning rate, $t(266) = .53$, $p = .590$, $d = .06$. Learning rate was positively correlated with age ($r = .44$, $p < .001$). We calculated positive and negative learning rate separately to investigate

possible differences across development. Both positive ($r = .42$, $p < .001$) and negative learning rate ($r = .34$, $p < .001$) correlated with age, but we did not find a correlation between age and the ratio between positive and negative learning rate.

We also performed these analyses with categorical age groups to pinpoint the exact ages of change. An ANOVA with Age group as between-subject variable and Learning Rate as dependent variable showed that learning rate improved with age (see Figure 2), $F(9, 258) = 11.13$, $p < .001$, $\eta^2 = .28$. LSD post hoc tests indicated that ages 8/9, 10, 11, and 13 performed poorer than the adult group (all $p < .05$). An additional ANOVA including only age groups 8/9 to 13 confirmed a steep increase in learning rate in this age range, $F(4, 140) = 7.34$, $p < .001$, $\eta^2 = .17$, and an analysis including age groups 14 to 18–25 confirmed no additional development in this age range, $F(4, 118) = 1.83$, $p = .129$, $\eta^2 = .06$.

A regression analysis was performed with Age (continuous) and Puberty (PDS, testosterone, and estradiol) as predictors for learning rate for boys and girls separately. We tested whether puberty explained additional variance above age by using the Enter method, with age as first predictor and puberty as second predictor. The model with puberty as a second predictor was a significantly better predictor in males than the model with age alone, with PDS as the only significant puberty predictor (Step 1: $R^2 = .29$; age: $\beta = .54$, $p < .001$; Step 2: $\Delta R^2 = .08$; age: $\beta = .76$, $p < .001$; PDS: $\beta = -.41$, $p = .002$). No such effect was found for testosterone ($\beta = -.06$, $p = .680$) or estradiol ($\beta = .06$, $p = .570$). In girls, puberty (PDS, testosterone, or estradiol) did not explain additional variance.

fMRI Results

Whole-brain Analysis

Learning versus application. To examine which areas were important for feedback learning, we calculated the

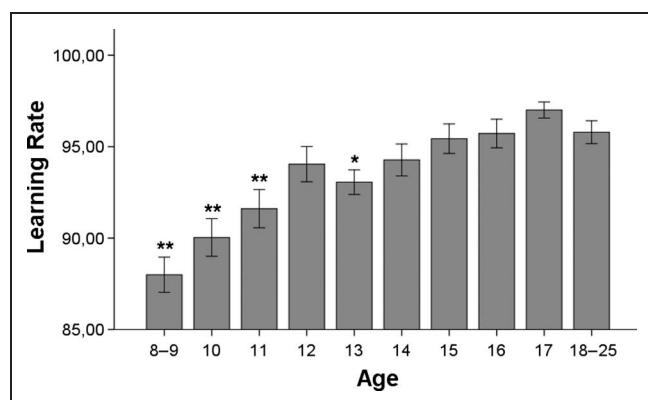
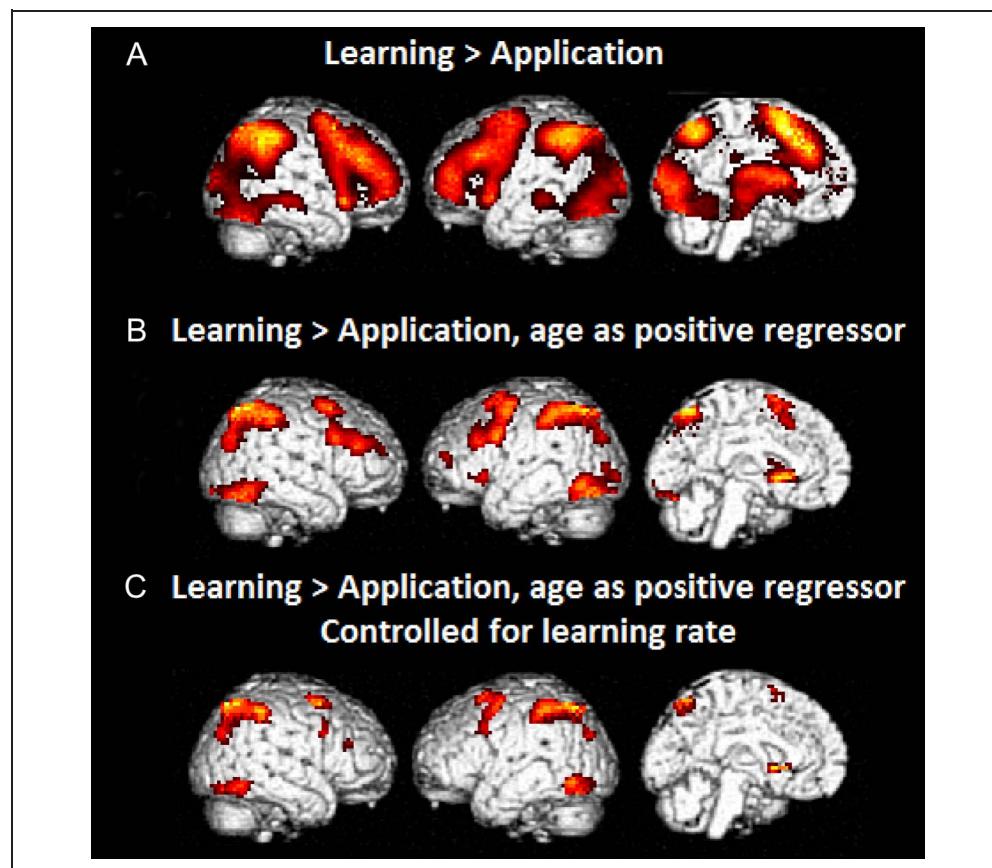


Figure 2. Learning rate per age group ($\pm SEM$). Asterisks represent a difference with the adult group, with one asterisk (*) indicating $p < .05$ and two asterisks (**) indicating $p < .01$.

Figure 3. (A) Learning (positive and negative) > Application. (B) Learning > Application, with age as a positive linear regressor. (C) Learning > Application, with age as a positive linear regressor, controlled for learning rate.



contrast Learning (positive and negative feedback combined) > Application. This contrast revealed widespread activation in the frontoparietal network including bilateral DLPFC, pre-SMA/ACC, and bilateral SPC (see Figure 3A; Table 2). Next, to test for developmental effects, we performed an analysis on the contrast Learning > Application with age as a continuous positive linear regressor, which resulted in activation in a similar but less widespread network (see Figure 3B; Table 2). We also tested for positive effects of age while controlling for learning rate to identify areas with specific sensitivity to increasing age. This resulted in largely overlapping activated areas compared with the analysis without performance control (see Figure 3C; Table 2). Finally, we performed an analysis on the contrast Learning > Application with learning rate as a positive regressor (controlled for age) to unravel activity patterns that were associated with better performance. We only found activity in a small cluster in left pre-SMA/ACC (local maximum at $-9, 15, 51, T = 5.23$, size = 13 voxels). The same analysis but with learning rate as a negative regressor did not result in any activation.

NegativeLearning versus PositiveLearning. The contrast NegativeLearning > PositiveLearning and the reverse contrast were calculated to identify areas sensitive to feedback valence during learning. NegativeLearning > PositiveLearning resulted in activity in bilateral DLPFC, pre-SMA/

ACC, anterior and mid-cingulum, SPC, and several subcortical areas (see Figure 4A; Table 3). Next, this analysis was performed with age entered as a positive linear regressor, which showed increased activity in bilateral SPC, bilateral DLPFC, and pre-SMA/ACC with increasing age (see Figure 4B; Table 3). This analysis was also performed with age as a positive regressor while controlling for learning rate to investigate activity that is specifically linked to age and not performance differences. This resulted in increased activity in superior frontal gyrus, SPC, and occipital areas (see Figure 4C; Table 3). Age as a negative regressor did not result in any activity. We also calculated this contrast with learning rate as a positive regressor (controlled for age), but this did not result in any significant activation. The same contrast, but with age as a negative regressor, resulted in activity in a small cluster in the right insula (local maximum at $30, -27, 24, T = 5.53$, size = 19 voxels).

PositiveLearning versus NegativeLearning. PositiveLearning > NegativeLearning revealed activation in bilateral precuneus, SPC, DLPFC, pre-SMA/ACC, and subcortical areas (see Figure 4D; Table 4). The analyses with age and learning rate as regressors were already covered in the previous paragraph on the contrast NegativeLearning > PositiveLearning.

Table 2. MNI Coordinates Local Maxima Activated for the Contrast Learning > Application

	<i>Area of Activation</i>	<i>x</i>	<i>y</i>	<i>z</i>	<i>Voxels</i>	<i>T</i>
<i>Learning > Application</i>						
Frontal cortex/subcortical	Superior medial frontal gyrus	0	24	42	11313	32.28
	R putamen	30	21	0		30.74
	R superior frontal gyrus	24	9	60		29.73
Parietal cortex	R inferior parietal lobule	45	-42	48	8653	29.86
	L inferior parietal lobule	-48	-45	48		25.92
	R precuneus	6	-66	48		24.35
Temporal cortex	L middle temporal gyrus	-57	-30	-9	113	12.77
	R middle cingulum	3	-30	27	12	5.94
<i>Learning > Application, Age as Positive Regressor</i>						
Frontal cortex	R middle frontal gyrus	30	9	60	521	9.29
	R inferior frontal gyrus	48	12	33		6.57
	R inferior frontal gyrus	45	33	24		6.29
	L precentral gyrus	-36	0	60	732	8.24
	L precentral gyrus	-48	3	51		7.68
	L superior frontal gyrus	-21	12	63		7.37
	L inferior frontal gyrus	-30	30	0	65	5.58
	L inferior frontal gyrus	-45	21	-3		5.42
	L middle frontal gyrus	-36	54	15	30	5.50
Subcortical	R caudate nucleus	6	21	0	260	7.00
	L caudate nucleus	-9	9	0		6.79
	R caudate nucleus	9	9	0		6.63
Parietal Cortex	R inferior parietal lobule	42	-39	48	2127	10.15
	R superior parietal lobule	21	-69	54		9.36
	L superior parietal lobule	-27	-63	54		9.07
Occipital Cortex	L inferior occipital gyrus	-48	-66	-15	407	8.54
	L fusiform gyrus	-33	-51	-18		5.54
	L middle occipital gyrus	-36	-84	6		5.24
Temporal cortex/occipital cortex/cerebellum	R inferior temporal gyrus	51	-60	-15	292	8.06
	R cerebellum	33	-69	-21		5.75
	R inferior occipital gyrus	27	-93	-12		4.96
	L cerebellum	-3	-81	-18	52	6.13
	L lingual gyrus	-12	-93	-15		4.89
<i>Learning > Application, Age as Positive Regressor, Controlled for Learning Rate</i>						
Frontal cortex	R middle frontal gyrus	30	12	60	76	7.56
	L precentral gyrus	-36	0	60	164	6.60
	L superior frontal gyrus	-21	12	63		6.19
	L precentral gyrus	-51	12	42		5.54

Table 2. (continued)

	<i>Area of Activation</i>	<i>x</i>	<i>y</i>	<i>z</i>	<i>Voxels</i>	<i>T</i>
Subcortical	R inferior frontal gyrus	51	12	33	25	5.02
	R middle frontal gyrus	51	12	45		4.79
	R inferior frontal gyrus	45	33	24	10	4.98
	R caudate nucleus	6	18	0	76	5.88
	L caudate nucleus	-6	9	0		5.70
	L caudate nucleus	-3	18	0		5.19
Parietal cortex/occipital cortex	R inferior parietal lobule	42	-39	48	626	8.16
	R superior parietal lobule	21	-69	54		7.21
	R middle occipital gyrus	33	-72	33		7.08
	L superior parietal lobule	-27	-63	54	480	6.99
	L inferior parietal lobule	-42	-45	57		6.50
	L inferior parietal lobule	-39	-45	48		6.34
	L inferior occipital gyrus	-48	-66	-15	123	6.75
	L fusiform gyrus	-33	-72	-18		5.05
	R inferior temporal gyrus	51	-60	-15	104	6.25
Temporal cortex/occipital cortex	R inferior occipital gyrus	42	-78	-15		5.52

Anatomical labels were acquired with automated anatomical labeling.

ROI Analysis: Valence Effects

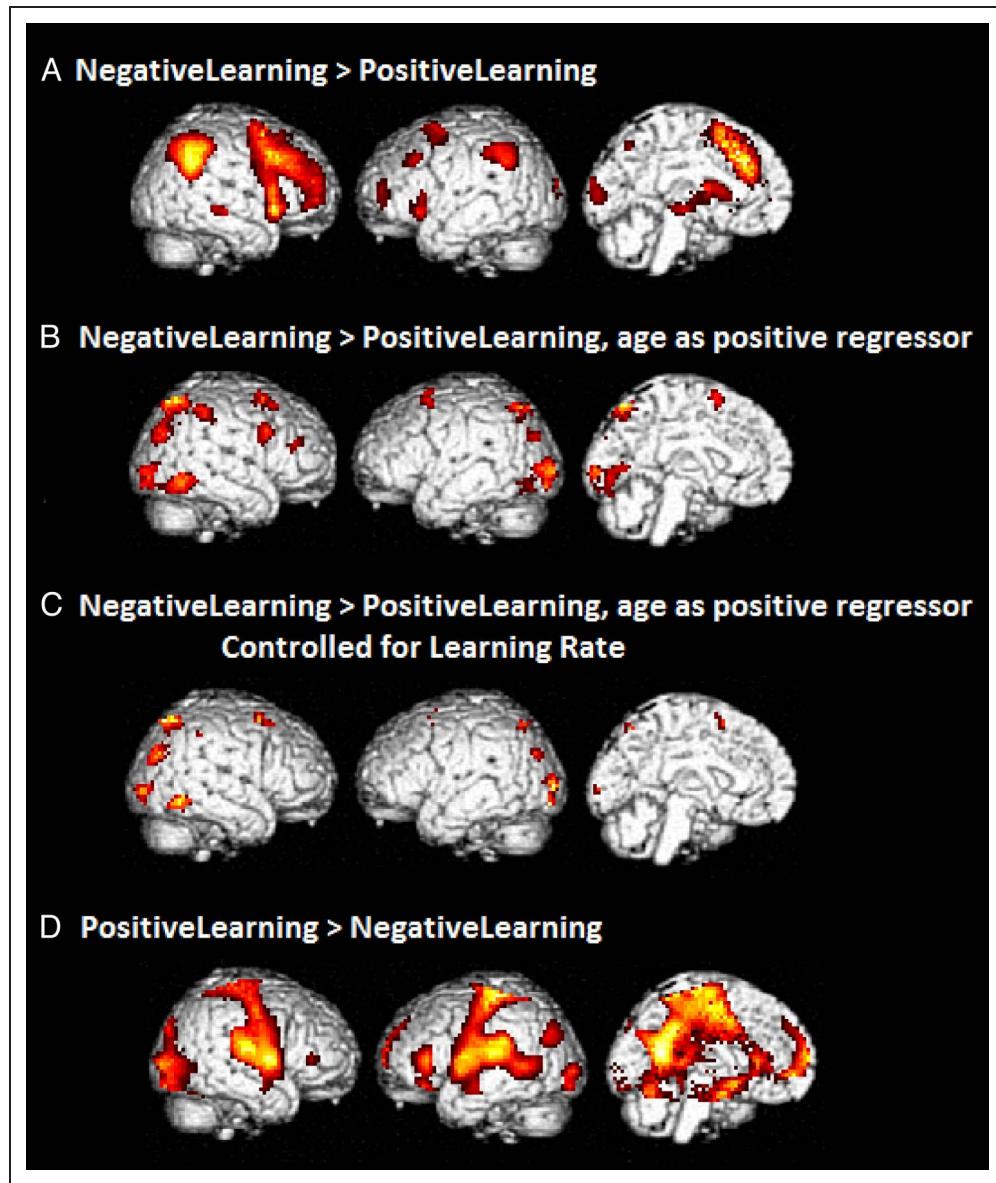
To pinpoint the exact ages at which changes in activation patterns occurred, we used ROI analyses. We focused on activity in pre-SMA/ACC, bilateral superior and mid-DLPFC, and bilateral SPC in relation to feedback valence, because prior studies found age differences for feedback learning in these areas (Crone et al., 2008; van Duijvenvoorde et al., 2008). Age was analyzed as a categorical age group to identify the precise time of change. The addition of sex did not result in any main or interaction effects. Therefore, effects of sex are not further discussed. In the following analyses, we will more specifically focus on valence effects and discuss them separately per region.

Right superior DLPFC. The Age \times Valence ANOVA for right superior DLPFC resulted in a main effect of Valence, $F(1, 258) = 244.02, p < .001, \eta^2 = .44$, and an Age \times Valence interaction, $F(9, 258) = 5.38, p < .001, \eta^2 = .09$ (see Figure 5). To follow up on this interaction, we tested for age differences in PositiveLearning and NegativeLearning separately. For NegativeLearning, there was a difference across age groups, $F(9, 258) = 5.36, p < .001, \eta^2 = .16$, but no age group effect was found for PositiveLearning, $F(9, 258) = 0.48, p = .886, \eta^2 = .02$. A second set of post hoc analyses tested for the valence effects within each age group. Paired-samples *t* tests were performed for

each age group between PositiveLearning and NegativeLearning. All ages except 8/9-year-olds showed a significant difference between NegativeLearning and PositiveLearning (all $p < .01$; see Figure 5). A third post hoc analysis tested at which the age the neural pattern for valence was adult-like. For this analysis, we calculated the difference between NegativeLearning and PositiveLearning, which differed across age groups, $F(9, 258) = 5.375, p < .001, \eta^2 = .16$. An LSD post hoc comparison with the adult group as baseline indicated that ages 8–13 differed significantly from the adult group (8/9 years: $p < .001$, 10 years: $p = .011$, 11 years: $p = .003$, 12 years: $p = .002$, 13 years: $p = .003$), whereas ages 14–17 did not differ significantly from adults.

Left superior DLPFC. The Age \times Valence ANOVA for left superior DLPFC showed a main effect of Valence, $F(1, 258) = 63.13, p < .001, \eta^2 = .18$, and an Age \times Valence interaction, $F(9, 258) = 3.38, p = .001, \eta^2 = .09$ (see Figure 5). To follow up on this interaction, we tested for age differences in PositiveLearning and NegativeLearning separately. There were no differences across age groups for NegativeLearning, $F(9, 258) = 1.78, p = .072, \eta^2 = .06$, and PositiveLearning, $F(9, 258) = 1.02, p = .421, \eta^2 = .03$. Further post hoc analyses tested for the valence effects within each age group. All ages from 14 to 18–25 showed a significant difference between NegativeLearning and PositiveLearning (all $p < .01$; see Figure 5). A final set of

Figure 4. (A) NegativeLearning > PositiveLearning. (B) NegativeLearning > PositiveLearning, with age as a positive linear regressor. (C) NegativeLearning > PositiveLearning, with age as a positive linear regressor, controlled for learning rate. (D) PositiveLearning > NegativeLearning.



post hoc analyses tested for the age at which the neural pattern for valence was adult-like. For this analysis, we used the difference score between NegativeLearning and PositiveLearning, which differed across age groups, $F(9, 258) = 3.38, p = .001, \eta^2 = .11$. An LSD post hoc comparison with the adult group as baseline indicated that ages 8/9, 11, 12, and 13 differed significantly from the adult group (8/9 years: $p = .006$, 11 years: $p = .006$, 12 years: $p = .018$, 13 years: $p = .021$), whereas ages 10 and 14–17 did not differ significantly from adults. The interaction between Valence \times Age \times Hemisphere was not significant, $F(9, 258) = .50, p = .872, \eta^2 < .001$, indicating there was no significant difference between left and right superior DLPFC.

Right mid-DLPFC. The Age \times Valence ANOVA for right mid-DLPFC resulted in a main effect of Valence, $F(1, 258) = 196.12, p < .001, \eta^2 = .41$, and an Age \times Valence

interaction, $F(9, 258) = 2.53, p = .009, \eta^2 = .05$. Follow-up tests indicated that NegativeLearning differed across age groups, $F(9, 258) = 2.21, p = .022, \eta^2 = .07$, but there was no age effect for PositiveLearning, $F(9, 258) = 1.47, p = .158, \eta^2 = .05$. Next, valence effects were tested for age groups separately. NegativeLearning resulted in stronger activations than PositiveLearning for all age groups (all $p < .05$; see Figure 5). Further post hoc tests were performed on differential activation for NegativeLearning compared with PositiveLearning (which differed across age groups: $F(9, 258) = 2.53, p = .009, \eta^2 = .08$) to test when activation patterns were adult-like. An LSD post hoc comparison indicated that none of the age groups differ significantly from the adult group.

Left mid-DLPFC. The Age \times Valence ANOVA for left mid-DLPFC resulted in a main effect of Valence, $F(1, 258) = 43.790, p < .001, \eta^2 = .14$, but no Age \times Valence

Table 3. MNI Coordinates for Local Maxima Activated for the Contrast NegativeLearning > PositiveLearning

	<i>Area of Activation</i>	<i>x</i>	<i>y</i>	<i>z</i>	<i>Voxels</i>	<i>T</i>
<i>NegativeLearning > PositiveLearning</i>						
Frontal cortex	R superior medial frontal gyrus	9	27	39	3638	17.88
	R insula	30	24	0		16.18
	R superior frontal gyrus	21	9	57		14.75
	L middle frontal gyrus	-42	24	39	60	6.80
	L middle frontal gyrus	-30	51	9	75	6.30
Frontal cortex/subcortical	L insula	-30	21	-3	552	11.23
	R caudate nucleus	15	18	9		9.42
	L caudate nucleus	-12	15	9		9.41
Parietal cortex	R angular gyrus	54	-51	33	923	15.25
	L inferior parietal lobule	-51	-54	39	303	8.47
	L inferior parietal lobule	-42	-42	42		7.73
	R precuneus	9	-63	48	38	7.15
	L precuneus	-9	-63	48	14	5.84
Temporal cortex	L calcarine gyrus	-9	-90	6	94	7.12
	R superior temporal gyrus	51	-24	-6	61	6.81
<i>NegativeLearning > PositiveLearning, Age as Positive Regressor</i>						
Frontal cortex	R superior frontal gyrus	24	9	57	150	6.52
	R inferior frontal gyrus	45	9	30	79	5.76
	L SMA	-12	6	66	69	5.65
	L superior frontal gyrus	-18	12	60		5.37
	L middle frontal gyrus	-30	6	57		5.04
	R middle frontal gyrus	48	39	21	23	5.08
Parietal cortex/occipital cortex	R superior parietal lobule	27	-63	54	891	7.31
	R middle occipital gyrus	30	-69	30		7.07
	R angular gyrus	27	-60	39		6.58
Temporal cortex/occipital cortex	R inferior temporal gyrus	51	-51	-12	265	6.28
	R inferior occipital gyrus	30	-90	-3		5.62
	R inferior occipital gyrus	36	-87	-12		5.30
Occipital cortex	L middle occipital gyrus	-24	-93	3	290	5.55
	L inferior occipital gyrus	-27	-93	-6		5.42
	L middle occipital gyrus	-30	-84	0		5.20
	L inferior occipital gyrus	-48	-75	-12	25	5.14
<i>NegativeLearning > PositiveLearning, Age as Positive Regressor, Controlled for Learning Rate</i>						
Frontal cortex	R superior frontal gyrus	21	9	51	96	6.31
	L superior frontal gyrus	-21	12	57	15	5.14
	L SMA	-12	6	66		4.94

Table 3. (continued)

	<i>Area of Activation</i>	<i>x</i>	<i>y</i>	<i>z</i>	Voxels	<i>T</i>
Parietal cortex	L superior parietal lobule	-21	-69	45	29	4.87
	L superior parietal lobule	-24	-69	54		4.86
	L superior parietal lobule	-15	-63	54		4.72
Parietal cortex/occipital cortex	R middle occipital gyrus	30	-66	30	306	6.24
	R superior parietal lobule	24	-63	54		5.94
	R superior occipital gyrus	24	-60	39		5.58
Temporal cortex	R inferior temporal gyrus	45	-57	-9	75	5.62
	R supramarginal gyrus	42	-42	42	13	5.00
Occipital cortex	L inferior occipital gyrus	-27	-93	-12	64	5.15
	L middle occipital gyrus	-27	-93	9		5.09
	L middle occipital gyrus	-30	-78	30	16	5.08
	R inferior occipital gyrus	27	-93	-3	50	5.04
	R middle occipital gyrus	33	-84	0		4.95
	R inferior occipital gyrus	36	-87	-12		4.82

interaction, $F(9, 258) = 1.78, p = .072, \eta^2 = .05$. Therefore, no further follow-up tests were performed. However, the Valence \times Age \times Hemisphere interaction was not significant, $F(9, 258) = .73, p = .684, \eta^2 = .002$, indicating that left and right mid-DLPFC showed a similar pattern of activation.

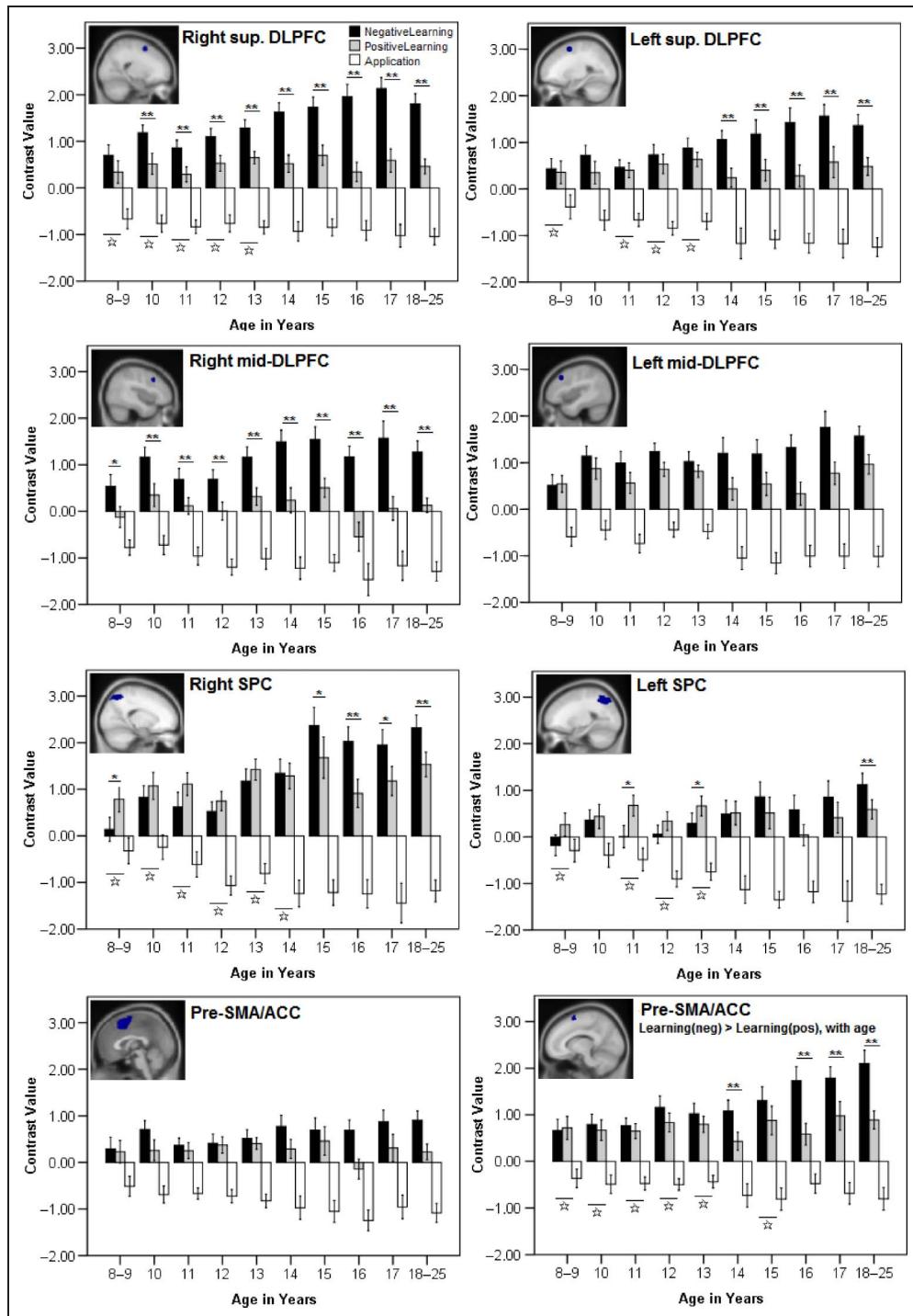
Right SPC. The Age \times Valence ANOVA for right SPC did not result in a main effect of Valence, $F(1, 258) = 3.32, p = .070, \eta^2 = .01$, but there was a significant Age \times Valence interaction, $F(9, 258) = 5.14, p < .001$,

$\eta^2 = .15$. Post hoc comparisons of PositiveLearning and NegativeLearning separately indicated that there was an effect of age group for NegativeLearning, $F(9, 258) = 7.56, p < .001, \eta^2 = .21$, but not for PositiveLearning, $F(9, 258) = 1.26, p = .262, \eta^2 = .04$. A further follow-up analysis for each age group separately indicated that ages 8/9, and 16 to 18–25 differentiated between PositiveLearning and NegativeLearning (all p s < .05). Notably, 8/9-year-olds show more activation after PositiveLearning, whereas ages 16 to 18–25 years show more activity after NegativeLearning (all p s < .05). Participants aged 10–14 showed

Table 4. MNI Coordinates Local Maxima Activated for the Contrast PositiveLearning > NegativeLearning

	<i>Area of Activation</i>	<i>x</i>	<i>y</i>	<i>z</i>	Voxels	<i>T</i>
<i>PositiveLearning > NegativeLearning</i>						
Frontal cortex/parietal cortex/subcortical	L precentral gyrus	-24	-27	66	14051	13.93
	L precuneus	-9	-57	12		13.11
	R caudate nucleus	21	-6	30		12.88
Frontal cortex	L medial orbital frontal gyrus	-3	60	-3	372	12.27
	L superior medial frontal gyrus	-9	66	12		9.96
	L superior medial frontal gyrus	-9	63	21		9.15
	R inferior frontal gyrus	54	33	6	21	7.91
Parietal cortex	L angular gyrus	-45	-75	27	117	7.87
Temporal cortex/occipital cortex	L middle occipital gyrus	-27	-93	0	132	8.51
	L inferior occipital gyrus	-33	-90	-9		7.16
	L lingual gyrus	-9	-93	-15		5.47

Figure 5. Activity for PositiveLearning, NegativeLearning, and Application in ROIs ($\pm SEM$). Asterisks represent a significant difference between PositiveLearning and NegativeLearning, with one asterisk (*) indicating $p < .05$ and two asterisks (**) indicating $p < .01$. Stars (*) indicate a significant difference with the adult group for difference scores between NegativeLearning and PositiveLearning.



no significant differentiation between PositiveLearning and NegativeLearning. A final post hoc test was performed to investigate when activation patterns reached adult levels. A one-way ANOVA indicated that differential activation for NegativeLearning compared with PositiveLearning differed across age groups, $F(9, 258) = 5.143, p < .001, \eta^2 = .15$. LSD post hoc comparisons indicated that ages 8–14 years differed significantly from the adult group (8/9 years: $p < .001$, 10 years: $p = .007$, 11 years: $p = .001$, 12 years: $p = .005$, 13 years: $p = .002$, 14 years: $p = .045$).

Left SPC. The analyses for left SPC also showed no main effect of Valence, $F(1, 258) < .001, p = .990, \eta^2 < .001$, but a significant Age \times Valence interaction, $F(9, 258) = 3.70, p < .001, \eta^2 = .11$. Follow-up tests indicated that NegativeLearning differed across age groups, $F(9, 258) = 2.78, p = .004, \eta^2 = .09$, but there was no age effect for PositiveLearning, $F(9, 258) = .58, p = .811, \eta^2 = .02$. NegativeLearning differed from PositiveLearning in ages 11, 13, and 18–25 (all $ps < .05$; see Figure 5), with importantly 11- and 13-year-olds showing more activity after

PositiveLearning, but 18- to 25-year-olds showing more activity after NegativeLearning. Further post hoc tests were performed on differential activation for NegativeLearning compared with PositiveLearning (which differed across age groups: $F(9, 258) = 3.70, p < .001, \eta^2 = .11$) to test when activation patterns reached adult levels. LSD post hoc comparisons indicated that ages 8/9 to 13 (except age 10, $p = .060$) differed significantly from the adult group (8/9 years: $p = .001$, 11 years: $p < .001$, 12 years: $p = .008$, 13 years: $p = .002$). The interaction between Valence \times Age \times Hemisphere was significant, $F(9, 258) = 2.58, p = .007, \eta^2 = .003$, showing that there was a significant difference between left and right SPC.

Pre-SMA/ACC. The Age \times Valence ANOVA for pre-SMA/ACC resulted in a main effect of Valence, $F(1, 258) = 28.43, p < .001, \eta^2 = .09$, but no significant Age \times Valence interaction, $F(9, 258) = 1.83, p = .063, \eta^2 = .05$, indicating that valence effects were present but did not differ per age group (see Figure 5). It was unexpected that no age effects were found in the pre-SMA/ACC, because age effects in this area were found in the whole-brain analyses PositiveLearning > NegativeLearning with age as a positive regressor. To further investigate age effects in the pre-SMA/ACC, we selected this whole brain activated cluster and masked it with an anatomical mask of the pre-SMA/ACC. Subsequently, we performed follow-up analyses on these ROI-extracted values to test which age groups were driving this effect. We first tested for age differences in PositiveLearning and NegativeLearning separately. For NegativeLearning, there was a difference across age groups, $F(9, 258) = 3.880, p < .001, \eta^2 = .11$, but no age group effect was found for PositiveLearning, $F(9, 258) = .523, p = .858, \eta^2 = .02$. A second set of post hoc analyses tested for valence effects within each age group. Paired-samples t tests were performed for each age group between PositiveLearning and NegativeLearning. Ages 14, 16, 17, and 18–25 showed a significant difference between NegativeLearning and PositiveLearning (all $p < .01$; see Figure 5). A third post hoc analysis tested for the age at which the neural pattern for valence was adult-like. For this analysis, we calculated the difference between NegativeLearning and PositiveLearning, which differed across age groups, $F(9, 258) = 3.956, p < .001, \eta^2 = .12$. An LSD post hoc comparison with the adult group as baseline indicated that ages 8/9 to 13, and 15 differed significantly from the adults (8/9 years: $p < .001$, 10 years: $p = .001$, 11 years: $p < .001$, 12 years: $p = .003$, 13 years: $p = .001$, 15 years: $p = .016$).

Effects of puberty on brain activation. To test for effects of puberty on brain activity, a regression analysis was performed with age as first predictor (continuous) and puberty (PDS, testosterone, and estradiol) as second predictor for the difference scores for NegativeLearning and PositiveLearning for boys and girls separately. PDS, testosterone, and estradiol were not significant additional contributors to neural activation.

Effects of structural brain development. To investigate whether structural brain development influenced the relation between age and brain activity, we performed a regression analysis with age as first predictor and cortical thickness as second predictor for the difference scores for NegativeLearning and PositiveLearning. We found an additional effect of cortical thickness above age in right SPC (Step 1: $R^2 = .10$; age: $\beta = .32, p < .001$; Step 2: $\Delta R^2 = .02$; age: $\beta = .36, p < .001$, cortical thickness: $\beta = .13, p = .042$), but not in other regions.

DISCUSSION

In this study, we investigated the development of feedback learning in a large sample of children, adolescents, and adults. The results indicate that, with increasing age, the frontoparietal network becomes increasingly more activated in response to feedback during learning compared with application of rules but contributes differentially to learning from positive and negative feedback across development. Specifically, we found that (a) activity in DLPFC, SPC, and pre-SMA/ACC after negative feedback increased with age, but activity for positive feedback remained constant; (b) activity in superior DLPFC, SPC, and pre-SMA/ACC reached adult levels around age 14/15; and (c) DLPFC, pre-SMA, and SPC contributed differently to learning in children, with SPC showing a shift in activity from positive to negative feedback. In the next paragraphs, these findings will be described in more detail.

Sensitivity to Negative Feedback Increases with Development

In adults, we found more activation following negative compared with positive feedback in DLPFC, pre-SMA/ACC, and SPC, consistent with prior studies (Zanolie et al., 2008; Holroyd et al., 2004). In childhood and early adolescence (ages 8–13/14), children developed into faster learners and showed increasing activation in superior DLPFC, SPC, and pre-SMA/ACC after negative feedback with increasing age. When controlling for learning performance, the more anterior portions of the DLPFC were no longer correlated with age, suggesting that these regions might be related more to performance than maturation per se. Intriguingly, in the SPC young children showed increased activation following positive feedback compared with negative feedback, a pattern that has also been observed in a prior study (van Duijvenvoorde et al., 2008). Also in this area, reactions to positive feedback did not change with development, whereas reactions to negative feedback increased with age. It therefore seems that this effect occurred mainly because of decreased activation after negative feedback in young children, rather than increased positive feedback compared with older participants. Together, these findings suggest that the frontoparietal network functions in a different way in children than in adults.

Transition to Adult-like Activity Patterns

One of the aims of this study was to investigate at which age this network functions in an adult-like way. In terms of behavior and neural activation, adolescents of age 14/15 years and older no longer differed from adults. These findings are the first to provide such a precise index of development, given that prior neuroimaging studies collapsed across wide age ranges (e.g., 13–17 years; van den Bos et al., 2009) or selected specific age groups (e.g., 11- to 13-year-olds vs. adults; van Duijvenvoorde et al., 2008). The transition to adult-like cognitive control functions is consistent with behavioral literature, which has suggested that cognitive development reaches adult levels in early to mid-adolescence (Huizinga, Dolan, & van der Molen, 2006; Luna, Garver, Urban, Lazar, & Sweeney, 2004).

One possible cause for immature activity in the frontoparietal network is immature structural brain development (Lu et al., 2009; Klingberg et al., 2002). In this study, we found preliminary evidence for a role of cortical thickness on activity patterns, such that right SPC cortical thickness explained additional variance in activity in addition to age. Future longitudinal studies are needed to confirm this finding. Additionally, developmental changes in white matter connectivity have been reported well into adolescence (Lebel & Beaulieu, 2011), which may also explain immature activity patterns in children. Other explanations come from resting state research. The network for cognitive control seems already in place in children, but the strength of connectivity within this network undergoes developmental changes (Jolles, van Buchem, Crone, & Rombouts, 2011). Also, the strength of short-range connections seems to decrease with age, whereas the strength of long-range connections increases with adolescent development (Dosenbach et al., 2010; Fair et al., 2009).

Another explanation for immature activity is that children differ from adults in strategy use. This is supported by findings that adults generally perform learning tasks in an efficient hypothesis-testing approach, whereas children are more likely to use an inefficient trial-and-error approach (Schmittmann, Visser, & Raijmakers, 2006). An intriguing question is whether there is a bias for positive feedback in young children. Possibly, it is adaptive for children to focus attention on positive feedback, because they do not have the capacity to use information derived from negative feedback during hypothesis testing (van Duijvenvoorde et al., 2008; Schmittmann et al., 2006).

Effects of Pubertal Development

Although this study provides evidence for a transition point in feedback processing at the age of 14/15, future research is needed to unravel the mechanisms behind this transition. One mechanism we explored was pubertal development. Theoretical models have suggested a dual processing network for adolescent development, such that the development of limbic areas is influenced

by puberty, whereas the functional development of the frontoparietal network occurs relatively independent of hormones (Steinberg et al., 2008; Nelson, Leibenluft, McClure, & Pine, 2005). Our findings are consistent with this model and show no additional influence of self-reported puberty stage, testosterone and estradiol on neural activity. Interestingly though, we did find an additional effect of pubertal development in boys on behavioral performance. However, given that no relation between puberty and neural activation in the frontoparietal network was found, the results support the assumption of the dual processing network that cortical development occurs relatively independently of pubertal development.

Limitations

This study has several limitations. First, we only analyzed the trials where learning from feedback was successful, and we have not investigated what happened in the small percentage of trials where learning did not occur. This would be an interesting direction for future research with a task that is more difficult and causes more errors during learning. Second, we only investigated the effects of testosterone, estradiol, and PDS in this study. Further research could focus on a more diverse array of puberty measures, such as having a physician assessing pubertal status or collecting multiple hormonal measurements on consecutive days. Finally, the current study was cross-sectional, which does not allow us to draw conclusions about change within individuals. Future longitudinal studies will be important to unravel within-person variance over time in performance, strategy use, and associated brain activity.

Conclusion

This study provides an overview of the development of feedback learning across adolescent development. Because of the large sample size, it was possible to pinpoint developmental trajectories across adolescence. This study is the first to show that activity in DLPFC, SPC, and pre-SMA/ACC after negative feedback increases with age until approximately age 14/15, after which adult levels are reached. We also demonstrated that the SPC shows a qualitative change in recruitment, with more activity in children after positive feedback, but more activity in late adolescents and adults after negative feedback. These findings are interpreted in terms of separable contributions of the frontoparietal network in childhood and more integrated function in adolescence and adulthood. These findings provide important starting points for searching for flexible periods for learning and eventually tailoring educational programs to the needs of children at different stages in development.

Acknowledgments

We thank all participants and their parents for their collaboration and everyone involved with the data collection for the

Braintime project. We thank Geoffrey Bird for his careful reading of the final manuscript. This work was supported by a grant from the European Research Council (ERC Innovative Ideas) awarded to E. A. C. (ERC-2010-StG-263234).

Reprint requests should be sent to Sabine Peters, Brain and Development Lab, Leiden University, Wassenaarseweg 52, 2333AK Leiden, The Netherlands, or via e-mail: s.peters@fsw.leidenuniv.nl.

REFERENCES

- Bennett, C. M., Wolford, G. L., & Miller, M. B. (2009). The principled control of false positives in neuroimaging. *Social, Cognitive and Affective Neuroscience*, 4, 417–422.
- Brett, M., Anton, J. L., Valabregue, R., & Poline, J. B. (2002). Region of interest analysis using an SPM toolbox. Paper presented at the 8th International Conference on Functional Mapping of the Human Brain, Sendai, Japan.
- Bunge, S. A., & Wright, S. B. (2007). Neurodevelopmental changes in working memory and cognitive control. *Current Opinion in Neurobiology*, 17, 243–250.
- Casey, B. J., Jones, R. M., & Somerville, L. H. (2011). Braking and accelerating of the adolescent brain. *Journal of Research on Adolescence*, 21, 21–33.
- Cococcia, C. A., Kollokian, V., Kwan, R. K. S., & Evans, A. C. (1997). Online interface to a 3D MRI simulated brain database. *Neuroimage*, 5, S425.
- Crone, E. A., & Dahl, R. E. (2012). Understanding adolescence as a period of social-affective engagement and goal flexibility. *Nature Reviews Neuroscience*, 13, 636–650.
- Crone, E. A., Wendelken, C., Donohue, S. E., & Bunge, S. A. (2006). Neural evidence for dissociable components of task-switching. *Cerebral Cortex*, 16, 475–486.
- Crone, E. A., Zanolie, K., Van Leijenhorst, L., Westenberg, P. M., & Rombouts, S. A. (2008). Neural mechanisms supporting flexible performance adjustment during development. *Cognitive, Affective and Behavioral Neuroscience*, 8, 165–177.
- Dale, A. M. (1999). Optimal experimental design for event-related fMRI. *Human Brain Mapping*, 8, 109–114.
- Dale, A. M., Fischl, B., & Sereno, M. I. (1999). Cortical surface-based analysis. I. Segmentation and surface reconstruction. *Neuroimage*, 9, 179–194.
- de Water, E., Braams, B. R., Crone, E. A., & Peper, J. S. (2013). Pubertal development and sex steroids are related to alcohol use in adolescents. *Hormones and Behavior*, 63, 392–397.
- Dosenbach, N. U. F., Nardos, B., Cohen, A. L., Fair, D. A., Power, J. D., Church, J. A., et al. (2010). Prediction of individual brain maturity using fmri. *Science*, 329, 1358–1361.
- Eliassen, J. C., Lamy, M., Allendorfer, J. B., Boespflug, E., Bullard, D. P., Smith, M. S., et al. (2012). Selective role for striatal and prefrontal regions in processing first trial feedback during single-trial associative learning. *Brain Research*, 1458, 56–66.
- Eppinger, B., Mock, B., & Kray, J. (2009). Developmental differences in learning and error processing: Evidence from ERPs. *Psychophysiology*, 46, 1043–1053.
- Fair, D. A., Cohen, A. L., Power, J. D., Dosenbach, N. U., Church, J. A., Miezin, F. M., et al. (2009). Functional brain networks develop from a “local to distributed” organization. *PLOS Computational Biology*, 5, e1000381.
- Fischl, B., & Dale, A. M. (2000). Measuring the thickness of the human cerebral cortex from magnetic resonance images. *Proceedings of the National Academy of Sciences, U.S.A.*, 97, 11050–11055.
- Fischl, B., Liu, A., & Dale, A. M. (2001). Automated manifold surgery: Constructing geometrically accurate and topologically correct models of the human cerebral cortex. *IEEE Transactions on Medical Imaging*, 20, 70–80.
- Fischl, B., Sereno, M. I., & Dale, A. M. (1999). Cortical surface-based analysis—II: Inflation, flattening, and a surface-based coordinate system. *Neuroimage*, 9, 195–207.
- Fischl, B., Sereno, M. I., Tootell, R. B. H., & Dale, A. M. (1999). High-resolution intersubject averaging and a coordinate system for the cortical surface. *Human Brain Mapping*, 8, 272–284.
- Forbes, E. E., & Dahl, R. E. (2010). Pubertal development and behavior: Hormonal activation of social and motivational tendencies. *Brain and Cognition*, 72, 66–72.
- Forman, S. D., Cohen, J. D., Fitzgerald, M., Eddy, W. F., Mintun, M. A., & Noll, D. C. (1995). Improved assessment of significant activation in functional magnetic resonance imaging (fMRI): Use of a cluster-size threshold. *Magnetic Resonance in Medicine*, 33, 636–647.
- Holroyd, C. B., & Coles, M. G. (2002). The neural basis of human error processing: Reinforcement learning, dopamine, and the error-related negativity. *Psychological Review*, 109, 679–709.
- Holroyd, C. B., Nieuwenhuis, S., Yeung, N., Nystrom, L., Mars, R. B., Coles, M. G. H., et al. (2004). Dorsal anterior cingulate cortex shows fMRI response to internal and external error signals. *Nature Neuroscience*, 7, 497–498.
- Huijzinga, M., Dolan, C. V., & van der Molen, M. W. (2006). Age-related change in executive function: Developmental trends and a latent variable analysis. *Neuropsychologia*, 44, 2017–2036.
- Jolles, D. D., van Buchem, M. A., Crone, E. A., & Rombouts, S. A. (2011). A comprehensive study of whole-brain functional connectivity in children and young adults. *Cerebral Cortex*, 21, 385–391.
- Klingberg, T., Forssberg, H., & Westerberg, H. (2002). Increased brain activity in frontal and parietal cortex underlies the development of visuospatial working memory capacity during childhood. *Journal of Cognitive Neuroscience*, 14, 1–10.
- Lebel, C., & Beaulieu, C. (2011). Longitudinal development of human brain wiring continues from childhood into adulthood. *Journal of Neuroscience*, 31, 10937–10947.
- Lu, L. H., Dapretto, M., O’Hare, E. D., Kan, E., McCourt, S. T., Thompson, P. M., et al. (2009). Relationships between brain activation and brain structure in normally developing children. *Cerebral Cortex*, 19, 2595–2604.
- Luna, B., Garver, K. E., Urban, T. A., Lazar, N. A., & Sweeney, J. A. (2004). Maturation of cognitive processes from late childhood to adulthood. *Child Development*, 75, 1357–1372.
- Nelson, E. E., Leibenluft, E., McClure, E. B., & Pine, D. S. (2005). The social re-orientation of adolescence: A neuroscience perspective on the process and its relation to psychopathology. *Psychological Medicine*, 35, 163–174.
- Peper, J. S., Mandl, R. C., Braams, B. R., de Water, E., Heijboer, A. C., Koolschijn, P. C., et al. (2012). Delay discounting and frontostriatal fiber tracts: A combined DTI and MTR study on impulsive choices in healthy young adults. *Cerebral Cortex*, 23, 1695–1702.
- Petersen, A. C., Crockett, L., Richards, M., & Boxer, A. (1988). A self-report measure of pubertal status—Reliability, validity, and initial norms. *Journal of Youth and Adolescence*, 17, 117–133.
- Pfeifer, J. H., & Allen, N. B. (2012). Arrested development? Reconsidering dual-systems models of brain function in adolescence and disorders. *Trends in Cognitive Sciences*, 16, 322–329.
- Raznahan, A., Shaw, P., Lalonde, F., Stockman, M., Wallace, G. L., Greenstein, D., et al. (2011). How does your cortex grow? *Journal of Neuroscience*, 31, 7174–7177.

- Schmittmann, V. D., Visser, I., & Raijmakers, M. E. J. (2006). Multiple learning modes in the development of performance on a rule-based category-learning task. *Neuropsychologia*, 44, 2079–2091.
- Schweinsburg, A. D., Nagel, B. J., & Tapert, S. F. (2005). fMRI reveals alteration of spatial working memory networks across adolescence. *Journal of the International Neuropsychological Society*, 11, 631–644.
- Shaw, P., Kabani, N. J., Lerch, J. P., Eckstrand, K., Lenroot, R., Gogtay, N., et al. (2008). Neurodevelopmental trajectories of the human cerebral cortex. *Journal of Neuroscience*, 28, 3586–3594.
- Somerville, L. H., Jones, R. M., & Casey, B. J. (2010). A time of change: Behavioral and neural correlates of adolescent sensitivity to appetitive and aversive environmental cues. *Brain and Cognition*, 72, 124–133.
- Steinberg, L., Albert, D., Cauffman, E., Banich, M., Graham, S., & Woolard, J. (2008). Age differences in sensation seeking and impulsivity as indexed by behavior and self-report: Evidence for a dual systems model. *Developmental Psychology*, 44, 1764–1778.
- Talairach, J., & Tournoux, P. (1988). *Co-planar stereotaxic atlas of the human brain*. New York: Thieme.
- Tamm, L., Menon, V., & Reiss, A. L. (2002). Maturation of brain function associated with response inhibition. *Journal of the American Academy of Child and Adolescent Psychiatry*, 41, 1231–1238.
- van den Bos, W., Guroglu, B., van den Bulk, B. G., Rombouts, S. A., & Crone, E. A. (2009). Better than expected or as bad as you thought? The neurocognitive development of probabilistic feedback processing. *Frontiers in Human Neuroscience*, 3, 52.
- van Duijvenvoorde, A. C., Zanolie, K., Rombouts, S. A., Raijmakers, M. E., & Crone, E. A. (2008). Evaluating the negative or valuing the positive? Neural mechanisms supporting feedback-based learning across development. *Journal of Neuroscience*, 28, 9495–9503.
- Welsh, M. C., Pennington, B. F., & Groisser, D. B. (1991). A normative developmental-study of executive function—A window on prefrontal function in children. *Developmental Neuropsychology*, 7, 131–149.
- Zanolie, K., Van Leijenhorst, L., Rombouts, S. A., & Crone, E. A. (2008). Separable neural mechanisms contribute to feedback processing in a rule-learning task. *Neuropsychologia*, 46, 117–126.